Montgomery County Community College 340 DeKalb Pike Blue Bell, PA 19422 Document Number: MET 14 Revision Number: 1 Effective Date: 10JAN14 Page 1 of 6

## **SOP: Eppendorf Research Plus Performance Verification**

#### **Approvals:**

| Preparer: Jason McMillan    | Date: 08JAN14 |
|-----------------------------|---------------|
| Reviewer: Dr. Maggie Bryans | Date: 10JAN14 |

- 1. Purpose: To verify the calibration of a single channel pipette.
- 2. Scope: Covers the cleaning, decontamination and verification of a single channel pipette.

#### 3. Responsibilities:

- 3.1. It is the responsibility of the course instructor/lab assistant to ensure that this SOP is performed as directed and to update the procedure when necessary.
- 3.2. It is the responsibility of the students/technicians to follow the SOP as described and to inform the instructor about any deviations or problems that may occur while performing the procedure.

#### 4. References:

- 4.1. balance operation SOP
- 4.2. balance calibration SOP
- 4.3. Tuttnauer 3850 ELV Autoclave SOP
- 4.4. Eppendorf Research Plus Operation and Maintenance SOP
- 5. **Definitions:** N/A
- 6. Precautions: N/A

#### 7. Materials:

- 7.1. balance
- 7.2. weigh boats
- 7.3. MilliQ water
- 7.4. small beaker for holding MilliQ water
- 7.5. verification labels
- 7.6. verification form
- 7.7. verification Pass/Fail form
- 7.8. pipette tips
- 7.9. Eppendorf Research Plus (P20, P200, and P1000)
- 7.10. 70% isopropyl alcohol (IPA)
- 7.11. lab towels
- 7.12. tweezers
- 7.13. thermometer
- 7.14. calculator
- 7.15. barometer

#### 8. Procedure:

8.1. Clean the pipette (See Figure 2.)

Note: Most pipettes are designed so that the parts that normally come into contact with liquid contaminants can easily be cleaned and decontaminated.

- 8.1.1. Wipe entire pipette with a lab towel dampened with a mild detergent solution.
- 8.1.2. Wipe entire pipette with a lab towel dampened with distilled water.
- 8.1.3. Remove the ejector sleeve by holding down the ejection button and pulling on the ejector sleeve (Figure 2: Step 1).
- 8.1.4. Slide up the ring on the lower part with the label "PUSH TO RELEASE"

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approximately 5mm until the lower part is released (Figure 2: Step 2 & 3).

- 8.1.5. The lower part is then removed from the upper part (Figure 2: Step 4).
- 8.1.6. Wipe the ejector sleeve and lower part with a lab towel dampened with a mild soap solution or 70% IPA.
- 8.1.7. Wipe the ejector sleeve and lower part with a lab towel dampened with MilliQ water.
- 8.1.8. Refit the lower part into the upper part until it engages audibly.
- 8.1.9. Refit the ejector sleeve and allow the pipette to dry.
- 8.1.10. Dispose of lab towels in bio-hazardous waste receptacle.

#### 8.2. Chemical decontamination

- 8.2.1. Spray a lab towel with 70% IPA to dampen the lab towel.
- 8.2.2. Wipe upper part of body with dampened lab towel.
- 8.2.3. Wipe ejector sleeve with dampened lab towel.
- 8.2.4. Wipe entire pipette with a lab towel dampened with MilliQ water.
- 8.2.5. Leave pipette to dry or wipe pipette dry with lab towel.
- 8.2.6. Dispose lab towels in bio-hazardous waste receptacle.

### 8.3. Autoclaving

- 8.3.1. Place the whole Eppendorf research Plus unit into the autoclave
- 8.3.2. Run the autoclave on the Unwrapped Delicate Instruments per Tuttnauer 3850 ELV Autoclave SOP
- 8.3.3. Remove the Eppendorf research Plus unit and allow it to dry completely and cool down.

#### 8.4. Verification of Calibration

Note: To test the accuracy of the pipette you will pipette a set volume 10 times and then weigh the total pipetted volume. 1mL of MilliQ water should weigh 1g and 1 $\mu$ L should weigh 1mg. Calculate your % Error using the equation below:

#### Expected Mass - Actual Mass x 100 = % Error Expected Mass

If the % Error is  $\leq 2\%$  the pipette passes verification, if the % Error is > than 2% the pipette fails. We will verify the pipette once at the maximum volume for the pipette, once at the  $\frac{1}{2}$  maximum volume, and once at the minimum volume. Altogether you will pipette 30 volumes and weigh 3 times for each pipette.

- 8.4.1. Record the necessary information on the Verification form. Enter information in the empty box to the right of the box specifying the information.
- 8.4.2. Verify that the calibration label of the balance is within the dated calibration time period.
- 8.4.3. Fill a small beaker with MilliQ water.
- 8.4.4. Place the weigh boat on the balance.
- 8.4.5. Tare the balance and verify that 0.00 is being displayed.
- 8.4.6. Verify that the pipette is set to the maximum volume (e.g. the maximum volume for a P-20 pipette is 20μL.).

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- 8.4.7. On the Pipette Verification Form, beside Selected Volume, enter the volume you will be pipetting and the value of that volume times 10 (e.g. for a  $20\mu$ L pipette you will record  $20\mu$ L for the selected volume and  $200\mu$ L for the selected volume times 10.).
- 8.4.8. Calculate the expected mass by converting the selected volume times 10 using the following conversions:  $1\mu L = 1mg$  and 1mL = 1g. Use the selected volume times 10 as the volume (e.g. for a  $20\mu L$  pipette,  $200\mu L$  multiplied by  $1mg/\mu L = 200mg$ ). Record the expected mass in the box beside Expected Mass.
- 8.4.9. Verify that the pipette is set to the maximum volume recommended by the manufacturer for the pipette.
- 8.4.10. Place pipette tip securely on the pipette.
- 8.4.11. Aspirate MilliQ water into pipette tip from the beaker and dispense it into weigh boat.
- 8.4.12. Repeat the above step 9 times. Each time you dispense the selected volume mark the Verification form in the numbered box beside Dispense Repetitions.
- 8.4.13. Record the final mass on the Verification form next to Recorded Mass.
- 8.4.14. Tare the balance and verify that 0.00 is being displayed.
- 8.4.15. Set the volume of the pipette to half capacity (e.g. For a P-20 pipette, set it to  $10\mu$ L.) and verify the volume.
- 8.4.16. Repeat steps 8.4.9. through 8.4.16 with the pipette set to the half-capacity volume.
- 8.4.17. Tare the balance and verify that 0.00 is being displayed.
- 8.4.18. Set the volume of the pipette to the minimum capacity recommended by the manufacturer (e.g. For a P-20 pipette, set it to 2μL.)
- 8.4.19. Repeat steps 8.4.9. through 8.4.16 with the pipette set to the minimum-capacity volume.
- 8.4.20. Calculate the % Error (as directed in the note at the beginning of section 8.4) for each test (maximum, half-capacity, and minimum volumes) and record the results on the verification form.
- 8.4.21. Verify that all fields of the verification form have been filled out and fill out the Pipette Verification Pass/Fail form according to the results of the tests.

#### 9. Attachments:

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**Figure 1: Eppendorf Research Plus** 



**Figure 2: Removing the Lower Part** Eppendorf Research Plus Manual

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## **10. History**

| Revision<br>Number | Effective<br>Date | Preparer       | Description of Change           |
|--------------------|-------------------|----------------|---------------------------------|
| 0                  | 08JAN14           | Jason McMillan | Initial release                 |
| 1                  | 10JAN14           | Jason McMillan | Added pipette verification form |

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| <b>Pipette Information</b> |  |  |  |  |  |  |  |
|----------------------------|--|--|--|--|--|--|--|
| Name and Description:      |  |  |  |  |  |  |  |
| Model:                     |  |  |  |  |  |  |  |
| Serial Number:             |  |  |  |  |  |  |  |

| Verification | Pipette            |  |
|--------------|--------------------|--|
| Technician   | Volume Range       |  |
| Date         | Number of Channels |  |

| Test  | Conditions |
|-------|------------|
| I COL | Conditions |

| Balance Serial #       | Balance Model            |
|------------------------|--------------------------|
| Sensitivity            | Balance Calibration Date |
|                        | Balance Calibration      |
| Correction Factor      | Technician               |
| Air Temperature        |                          |
| Barometric Temperature |                          |
| Relative Humidity      |                          |

| Tests                       |   |   |   |   |   |   |   |   |   |    |               |
|-----------------------------|---|---|---|---|---|---|---|---|---|----|---------------|
| Test 1 (Max. volume)        |   |   |   |   |   |   |   |   |   |    |               |
| Selected Volume             |   |   |   |   |   |   |   |   |   |    | Expected Mass |
| Selected Volume X 10        |   |   |   |   |   |   |   |   |   |    | Recorded Mass |
| <b>Dispense Repetitions</b> | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |               |
| Test 2 (Half cap. volume)   |   |   |   |   |   |   |   |   |   |    |               |
| Selected Volume             |   |   |   |   |   |   |   |   |   |    | Expected Mass |
| Selected Volume X 10        |   |   |   |   |   |   |   |   |   |    | Recorded Mass |
| Dispense Repetitions        | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |               |
| Test 3 (Min. volume)        |   |   |   |   |   |   |   |   |   |    |               |
| Selected Volume             |   |   |   |   |   |   |   |   |   |    | Expected Mass |
| Selected Volume X 10        |   |   |   |   |   |   |   |   |   |    | Recorded Mass |
| Dispense Repetitions        | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |               |

## Test results

| % Error Test 1 |  |
|----------------|--|
| % Error Test 2 |  |
| % Error Test 3 |  |
| Pass or Fail   |  |